



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>6</sup> : <b>A01N 63/00, A61K 7/48, 7/06, 7/16, 7/28</b>		<b>A1</b>	(11) International Publication Number: <b>WO 97/47202</b>
			(43) International Publication Date: 18 December 1997 (18.12.97)
(21) International Application Number: <b>PCT/EP97/03046</b>			(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).
(22) International Filing Date: 9 June 1997 (09.06.97)			
(30) Priority Data: 96201651.5 7 June 1996 (07.06.96) EP			
(34) Countries for which the regional or international application was filed: AT et al.			
(71) Applicant (for all designated States except US): GIST-BROCADES B.V. [NL/NL]; Wateringseweg 1, P.O. Box 1, NL-2600 MA Delft (NL).			
(72) Inventor; and (75) Inventor/Applicant (for US only): BEUDEKER, Robert, Franciscus [NL/NL]; Boomkwekerij 31, NL-2635 KC Den Hoom (NL).			
(74) Agents: VISSER-LUIRINK, Gesina et al.; Gist-Brocades N.V., Patents and Trademarks Dept., Wateringseweg 1, P.O. Box 1, NL-2600 MA Delft (NL).			Published With international search report.

(54) Title: ANTIFUNGAL COMPOSITIONS

## (57) Abstract

The invention provides synergistic combinations of fungicides and enzymes breaking down cell walls of yeasts and filamentous fungi. The fungicides are polyene macrolides, preferably nystatin and/or natamycin. The cell wall degrading enzymes are preferably used in combinations. They include cellulases, chitinases, mannanases, proteases and the like. The use of compositions according to the invention in the field of crop protection, especially for protection of flower bulbs, is also disclosed.

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### Antifungal compositions

5           The present invention relates to compositions for combatting (or killing) yeasts and filamentous fungi, herein referred to as fungicides or fungicidal compositions.

Fungicides are often used in crop protection, disinfection, cleaning and even cosmetics and pharmaceuticals.

10           In many of these uses the fungicides place an undesirable burden on the environment. However, fungus related diseases result in reductions of crop yields and present a health hazard for animals and humans due to the production of mycotoxins which may enter the food chain.

15           The cell wall of fungi is composed of carbohydrates such as chitin, glucan and mannan. Chitin is a polysaccharide composed of N-acetyl-D-glucosamine linked by  $\beta(1\rightarrow4)$ -glucosidic linkages. Mannans are composed of D-mannan, linked in the b-configuration by  $\beta(1\rightarrow4)$ -,  $\beta(1\rightarrow6)$ - and  
20  $\beta(1\rightarrow3)$ -linkages.  $\beta$ -glucans are homopolymers of D-glucose linked in the  $\beta$ -configuration. The occurrence and relative importance of these carbohydrates varies between classes of fungi. Oomycetes, for example, are characterized by a lack of chitin in the cell wall but do contain glucan and mannan.

25           Enzymes have been characterized which are capable of degrading fungal cell walls. These enzymes comprise endochitinases (EC 3.2.1.14), which randomly cleave chitin; chitobiosidases (chitin 1,4- $\beta$ -chitobiosidase; EC 3.2.1.30) which cleave dimeric units from one end of chitin; 1,3-  
30  $\beta$ -glucanases (glucan-1,3- $\beta$ -glucosidase; EC 3.2.1.58), which cleave 1,3- $\beta$ -glucans; and glucosaminidase (N-acetyl- $\beta$ -D-glucosaminidase; EC 3.2.1.30), which cleave monomeric units from one end of chitin and have N-acetyl- $\beta$ -glucosaminidase activity.

35           Several classes of micro-organisms secrete these enzymes into their environment. In particular, enzymes produced by the fungus *Trichoderma harzianum* and *Gliocladium virens* have been studied in detail. Genes encoding these enzymes have been cloned and the antifungal activities of

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these enzymes have been tested against plant pathogenic fungi. A variety of synergistic antifungal effects between two or more single purified enzymes have been demonstrated (Lorito, M., Hayes, C.K., DiPietro, A., Woo, S.L. and  
5 Harman, G.E., *Phytopathology* 84, 398-405 (1994)).

Combinations of chitinolytic and glucanolytic enzymes have been shown to be very effective in improving the fungicidal activity of the fungicides gliotoxin, flusilazole, miconazole, captan and benomyl thereby allowing  
10 a significant reduction in the chemical doses required (Lorito, M., Hayes, C.K., Zoina, A., Scala, F., Del Sorbo, G., Woo, S.L. and Harman, G.E., *Molecular Biotechnology* 2: 209-217 (1994)).

International Patent Application WO94/13784 describes  
15 combinations of fungal cell wall degrading enzymes and specific fungicides such as flusilazole, miconazole and captan.

The experiments described in the prior art have been conducted on a very small scale in the lab by using *in vitro*  
20 antifungal bioassays as described by Lorito, M., Harman, G.E., Hayes, C.K., Broadway, R.M., Tronsmo, A., Woo, S.L. and DiPietro, A., *Phytopathology* 83, 302-307 (1993). However, it is common that results in the lab cannot be applied to practical situations.

Moreover, the fungicides disclosed in the above mentioned patent application are all either sterol synthesis  
25 inhibitors or thiol group inactivators.

The present invention thus provides a fungicidal composition comprising a polyene macrolide antibiotic  
30 together with at least one fungal cell wall degrading enzyme.

Polyene macrolide antibiotics to be used in the compositions according to the invention include compounds such as pimarycin (natamycin) and nystatin. They are rarely, if  
35 ever, used in crop protection because of their high price, but are used in, for example, pharmaceutical fungicidal compositions.

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The combination of the macrolide and the cell wall degrading enzyme results in significant reduction in the amount of fungicide required so that the expensive macrolides can be applied as crop protection agents.

5 In addition, the increased efficacy of the compositions according to the invention over the separate components makes them very useful in other antifungal applications.

The polyene macrolide antifungal substances to be used  
10 in the composition according to the invention include, but are not limited to nystatin, natamycin, amphotericin B, candicidin, filipin, homycin, etruscomycin and trichomycin. Some of these macrolide compounds are mixtures of different active substances. These polyene macrolide antibiotics are  
15 characterized by a macrolide ring. They differ in the number (12-37) of carbon atoms in the ring structure, the number of hydroxyl groups (6-14) and the presence or absence of a carbohydrate. Polyene macrolide antibiotics alter the membrane permeability of fungal cells by forming a complex with  
20 sterol. As a result a fatal loss of potassium occurs.

The preferred fungicides according to the invention are natamycin and/or nystatin. These are very effective fungicides which show synergistic antifungal action in combination with cell wall degrading enzymes, particularly  
25 with  $\beta$ -1,3-glucanase, which is therefore a preferred cell wall degrading enzyme for use in the compositions according to the invention. However,  $\beta$ -1,3-glucanase may also break down plant cell walls and therefore the amount of this enzyme should be limited to 500,000 U/l (or kg), preferably  
30 50,000 U/l (or kg), more preferably 10,000 U/l (or kg). Its effectiveness, even in small amounts, can be maintained by using a combination of different cell wall degrading enzymes, such as a combination of  $\beta$ -1,3-glucanase and an enzyme which breaks down the components of fungal cell walls  
35 not present in plant cell walls. Such enzymes are for instance chitinase or mannanase. However, other enzymes may also be used, either alone or in combination. Useful enzymes, some of which have already been mentioned, include

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but are not limited to: cellulases, in particular exo/endoglucanases, such as  $\beta$ -1,3-glucanase or  $\beta$ -1,4-glucanase; exo/endochitinases; mannanases; galactanases and proteases.

5 The enzymes may be obtained from any organism which produces them. The exemplified enzymes have been obtained from *Trichoderma longibrachiatum*, but other micro-organisms are a source of enzymes as are plant cells, yeast cells, fungi and even animal or insect cells. It is of course clear  
10 that genes encoding cell wall degrading enzymes may be incorporated into any suitable host cell to facilitate production of the enzymes. Many useful enzymes have been disclosed on pages 4-12 of WO94/13784 which is incorporated herein by reference.

15 There are many areas in which the compositions according to the invention can be used. The nature of the antifungal compositions is determined by the use and is dependent on, for example, the manner of application and the effective dose.

20 Preferred areas of application include but are not limited to: antifungal treatment of seeds, bulbs, fruits, plants, silage, food, feed or fodder and the use in, for example, cosmetics and cleaning agents. Calculation of the required dosages may be performed by any person skilled in  
25 the art.

Compositions according to the invention will typically contain between 1 and 1000 mg/l of fungicide and between 50 and 500,000 Units of each enzyme/l (or kg), preferably between 1 and 500 mg/l fungicide and between 50 and 50,000  
30 Units of each enzyme/l (or kg), and most preferably between 1 and 250 mg/l fungicide and between 50 and 10,000 Units of each enzyme/l (or kg).

The compositions according to the invention can be used in essentially the same way as prior art antifungal  
35 compositions.

For agricultural applications, the compositions can be typically applied to seeds, roots, foliage or fruit. The

preferred agricultural products to be treated with the compositions according to the invention are flower bulbs.

For flower bulbs it is common to treat them with hot-water prior to planting to control parasitic insects, mites and nematodes. Such a treatment may prevent the spread of pathogenic micro-organisms (Langerak, 1985; PhD thesis entitled "The pathogenesis of *Fusarium oxysporium* f.sp. *nacissi* and the role of antagonistic bulb-borne fungi in the chemical control of basal rot" Agricultural University Wageningen, The Netherlands).

Antimicrobial agents may be added to the hot-water bath to reduce the spread of micro-organisms. The potential use of the antifungal agent natamycin (a polyene macrolide produced by *Streptomyces natalensis*) in this application has been demonstrated in the past (Langerak supra). Practical applications before now have been very limited due to the fact that other antifungal agents appeared to show a superior price: performance ratio. However, the effective dose rate of natamycin (or other fungicidal polyene macrolides) may be lowered considerably if sufficient units of the fungal cell wall degrading enzymes chitinase and  $\beta$ -1,3-glucanase are added according to the present invention. This finding improves the price performance ratio of natamycin considerably and enables commercial application.

In addition, mixing of fungicidal polyene macrolides such as natamycin and fungal cell wall degrading enzymes in the soil is a very effective way to control infection of flower bulbs by *Rhizoctonia*. Synergistic effects of fungicidal polyene macrolides such as natamycin and fungal cell wall degrading enzymes such as chitinase and  $\beta$ -1,3-glucanase result in the decrease of the necessary dose and can improve the price: performance ratio significantly.

A composition according to the invention can thus be added to the said hot-water bath, or the composition may be used as the hot aqueous bath itself.

The following examples are designed to illustrate and in no way limit the scope of the present invention.

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Example 1Production of fungal cell wall degrading enzymes

Enzymes were derived from a commercial fermentation of  
5 the fungus *Trichoderma longibrachiatum*. Broth of  
*T. longibrachiatum* was subjected to plate filtration fol-  
lowed by ultrafiltration. The ultrafiltrate was then sub-  
jected to fluid bed granulation according to procedures  
known to persons skilled in the art. Sodium sulphate was  
10 used as a nucleus during fluid bed granulation.

The tested preparation contained 180 units of  
endochitinase activity per gram of granulated product. One  
unit is defined as the amount of enzyme that liberates 1  $\mu$ -  
N-acetyl-D-glucosamine from cm-chitin-rbv (Sigma catalogue  
15 number C 3020) at pH 6.0 at 25°C per 48 hours.

The method has been described in detail in Analytical  
Biochemistry 8: 397-401 (1964). The same preparation also  
contained 50 units of  $\beta$ -1,3-glucanase per gram of granulated  
product. One unit is defined as the amount of enzyme that  
20 liberates 1 micromole of reducing sugars per minute from  
0.1% (w/v) laminarin (Sigma catalogue number L9634) at pH  
6.7 at 30°C. This method has been described in detail in  
Molecular Biotechnology 2: 209-217 (1994).

Example 2Synergistic effects of fungal cell wall degrading enzymes  
and natamycin against Fusarium in flower bulbs

25 Viability of conidia of *Fusarium oxysporum* was  
measured at the start and at the end of the hot-water treat-  
ment. (2 hrs at 43.5°C).

When no fungicidal agents were added, duplicate samples of  
5 ml were taken from the water in the bath at the beginning  
and the end of the treatment, and diluted with sterile water  
35 so that 1 ml did not contain more than 200 living conidia.  
Five samples of 1 ml were mixed each with 20 ml of PDA  
containing 50 microgram/ml vendarcin (PDA-V) and poured into



petri dishes of 14 cm diameter. Colonies were counted after 2 and 5 days at 25°C.

In the presence of natamycin and fungal cell wall degrading enzymes, 2 samples of 25 ml were taken from the bath at the beginning and the end of the treatment. These samples were centrifuged at 3,000 \* g for 15 min. The pellet containing the conidia was washed several times with sterile water to remove the antifungal agents, followed by centrifugation and dilution to allow for plating on PDA-V as described above.

Natamycin was added to the hot-water bath to a final concentration of 300 mg/L in the absence of fungal cell wall degrading enzymes. A series of natamycin concentrations were tested to demonstrate synergy with chitinase and  $\beta$ -1,3-glucanase.

Chitinase and  $\beta$ -1,3-glucanase respectively were added to the hot-water bath to a final activity of 36,000 and 10,000 units per liter. Units are as defined in Example 1. Results are shown in Table 1.

Table 1

Effects on survival of hot-water treated conidia (25,000 conidia per ml) of *Fusarium oxysporum* as measured on PDA-V agar. Fungal cell wall degrading enzymes and different concentrations of natamycin were added to the hot-water bath

Treatment	Surviving conidia per ml
No addition	100
Natamycin 300 mg/L	<1
Natamycin 200 mg/L	50
Natamycin 100 mg/L	70
Natamycin 100 mg/L plus fungal cell wall degrading enzymes	<1

**Example 3****Synergistic effects of fungal cell wall degrading enzymes and natamycin in the soil against Rhizoctonia in flower bulbs**

5

Bulbs were planted in pots at standard soil at a depth of 20 cm. Oat borne sclerotia of *Rhizoctonia* were mixed through the soil to infect the bulbs. Temperature was maintained at 18°C during the experiment. Experiments were replicated 5 fold.

10

Natamycin and fungal cell wall degrading enzymes were mixed as powders through the soil prior to infection with *Rhizoctonia*.

15

Natamycin was tested at final concentrations of 1000, 500 and 100 mg/kg. Chitinase and  $\beta$ -1,3-glucanase was mixed through the soil to a final activity of 3600 and 1000 units per kg of soil.

After 4 weeks bulbs were monitored for fungal infection by means of visual inspection. Results are shown in Table 2.

20

**Table 2**

Treatment	Percentage of bulbs showing fungal infection
No treatment	61
Natamycin 1000 mg/kg	19
Natamycin 500 mg/kg	42
Natamycin 100 mg/kg	50
Natamycin 100 mg/kg plus fungal cell wall degrading enzymes	21

30

35

The results demonstrate the synergistic effects of natamycin and fungal cell wall degrading enzymes on the plant pathogenic fungus *Rhizoctonia*. Differences between the

various treatments are statistically significant except for the groups "no treatment" and natamycin at a dose of 100 mg/kg.

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Claims

1. A fungicidal composition comprising a polyene  
5 macrolide antibiotic and at least one fungal cell wall  
degrading enzyme.

2. A composition according to claim 1 wherein the  
polyene antibiotic is natamycin or nystatin.

10

3. A composition according to claim 1 or 2 wherein  
the cell wall degrading enzyme comprises a cellulase, e.g. a  
glucanase, a mannanase, a chitinase, a galactanase or a  
protease.

15

4. A composition according to claim 3 wherein the  
glucanase is  $\beta$ -1,3-glucanase or a  $\beta$ -1,4-glucanase.

5. A composition according to claim 3 or 4 wherein  
20 the composition comprises a glucanase together with a  
chitinase or a mannanase.

6. A method for preparing of a composition according  
to any one of the proceeding claims which comprises mixing  
25 the fungicidal polyene antibiotic with at least one fungal  
cell wall degrading enzyme.

7. Use of a composition according to anyone of  
claims 1 - 5 in an antifungal treatment.

30

8. Use according to claim 7 in the treatment of  
seeds, bulbs, fruits, plants, silage, food, feed or fodder

9. Use according to claim 7 in a disinfectant or in  
35 cosmetics.

10. Use according to claim 8 in the treatment of seed  
potatoes.

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11. A composition according to anyone of claims 1-5 which is a detergent, disinfectant, cosmetic or agricultural composition.

5 12. A composition according to claim 11 which additionally comprises water and/or a detergent.

13. A method of treating a plant or part thereof, the method comprising contacting the plant or part thereof with  
10 a composition as defined in anyone of claims 1 to 5.

14. A method according to claim 13 wherein the part of a plant is a seed, tuber, bulb or fruit.

15 15. A part of a plant treated by a method according to claim 14.

## INTERNATIONAL SEARCH REPORT

Intern. Application No.  
PC1/EP 97/03046

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 A01N63/00 A61K7/48 A61K7/06 A61K7/16 A61K7/28

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A01N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	FR 2 385 730 A (SEARLE & CO) 27 October 1978 see claims 1,3-6 see example 3 ---	1-7,9, 11,12
X	EP 0 702 897 A (KAO CORPORATION) 27 March 1996 see page 2, line 40 - line 46 see page 2, line 51 - line 54 see page 3, line 5 - line 8 see page 7, line 10 - line 29 see claim 1 -----	1-8, 10-15

☐ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

21 August 1997

Date of mailing of the international search report

28 -08- 1997

Name and mailing address of the ISA

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NL - 2280 HV Rijswijk  
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Information on patent family members

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Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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